

Supplementary material of the manuscript published in Vitis **60**, 69–75 (2021):

Comparison of two sample preparation methods for ^1H -NMR wine profiling: Direct analysis and solid-phase extraction

I. OCAÑA-RIOS¹⁾, F. RUIZ-TERÁN²⁾, M. E. GARCÍA-AGUILERA¹⁾, K. TOVAR-OSORIO¹⁾, E. RODRÍGUEZ DE SAN MIGUEL²⁾ and N. ESTURAU-ESCOFET¹⁾

¹⁾Instituto de Química, Universidad Nacional Autónoma de México, Ciudad de México, México

²⁾Facultad de Química, Universidad Nacional Autónoma de México, Ciudad de México, México

Supplementary Fig. S1-S3: ^1H -NMR expansion spectra (700 MHz, D_2O , 25 °C) of eight wine samples by DA-NMR.

Supplementary Fig. S4-S6: ^1H -NMR expansion spectra (700 MHz, $\text{MeOD}-d_4$, 25 °C) of eight wine samples by SPE-NMR.

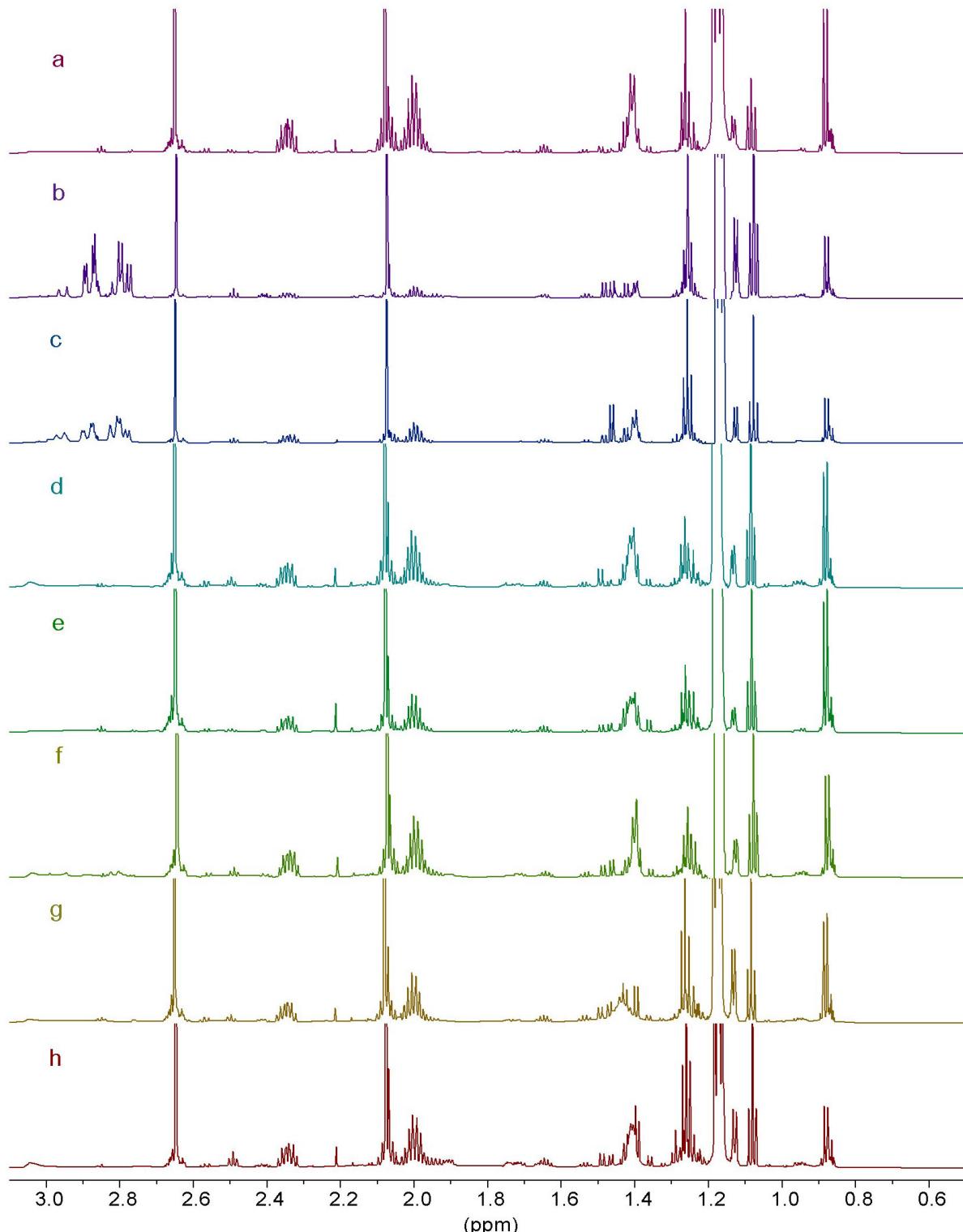
Supplementary Fig. S7: ^1H -RMN spectra (700 MHz, $\text{MeOD}-d_4$, 25 °C) of triplicate methanol extracts of a 'Cabernet Sauvignon' wine sample performed to evaluate SPE-NMR repeatability.

Supplementary Fig. S8: Control chart of the first component PC1 that explains 94 % of the data variability. Cliff plot showing the explained variance of the PCA model with a principal component.

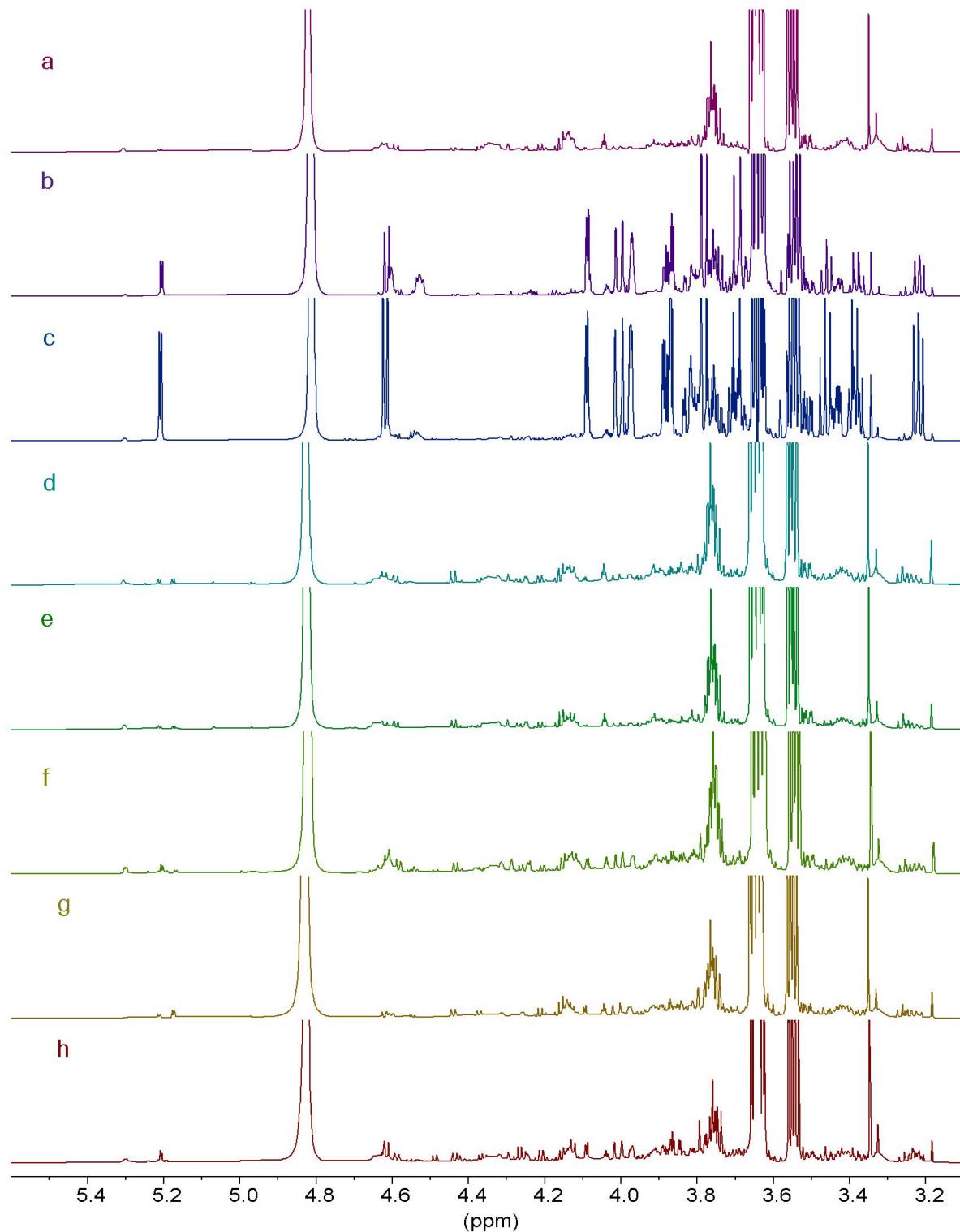
Supplementary Fig. S9-S10: Overlapped ^1H -NMR spectra of phenethyl and isoamyl alcohol signals in the eight wine samples by DA-NMR and SPE-NMR.

Supplementary work flow S11: Structural elucidation of tyrosol using 1D and 2D NMR spectra in 'Merlot' wine.

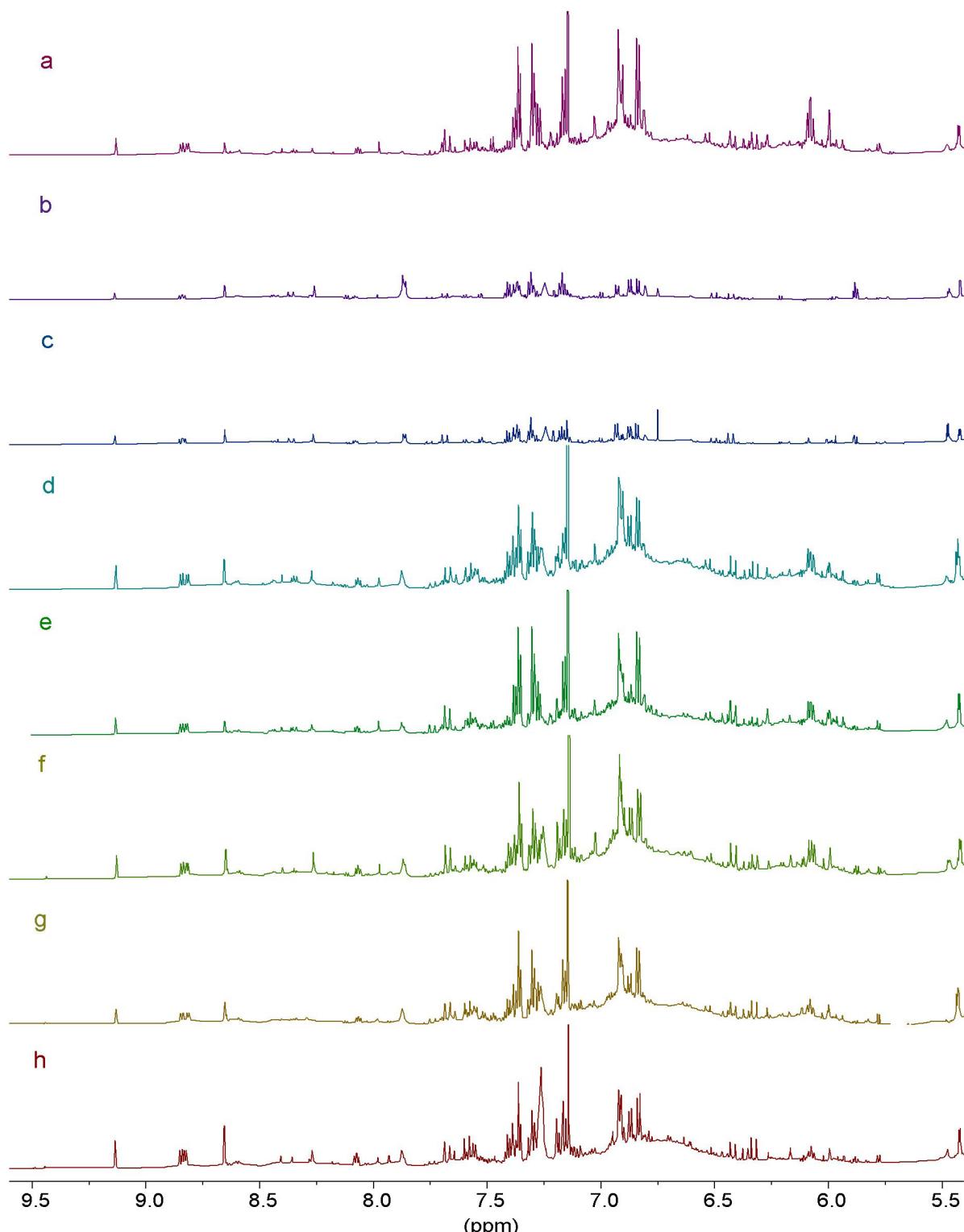
Supplementary Fig. S12-S16: ^1H , ^{13}C , COSY, ed-HSQC and HMBC spectra (700 MHz, $\text{MeOD}-d_4$, 25 °C) of 'Merlot' wine extracted by SPE-NMR. Signal assignments of tyrosol.



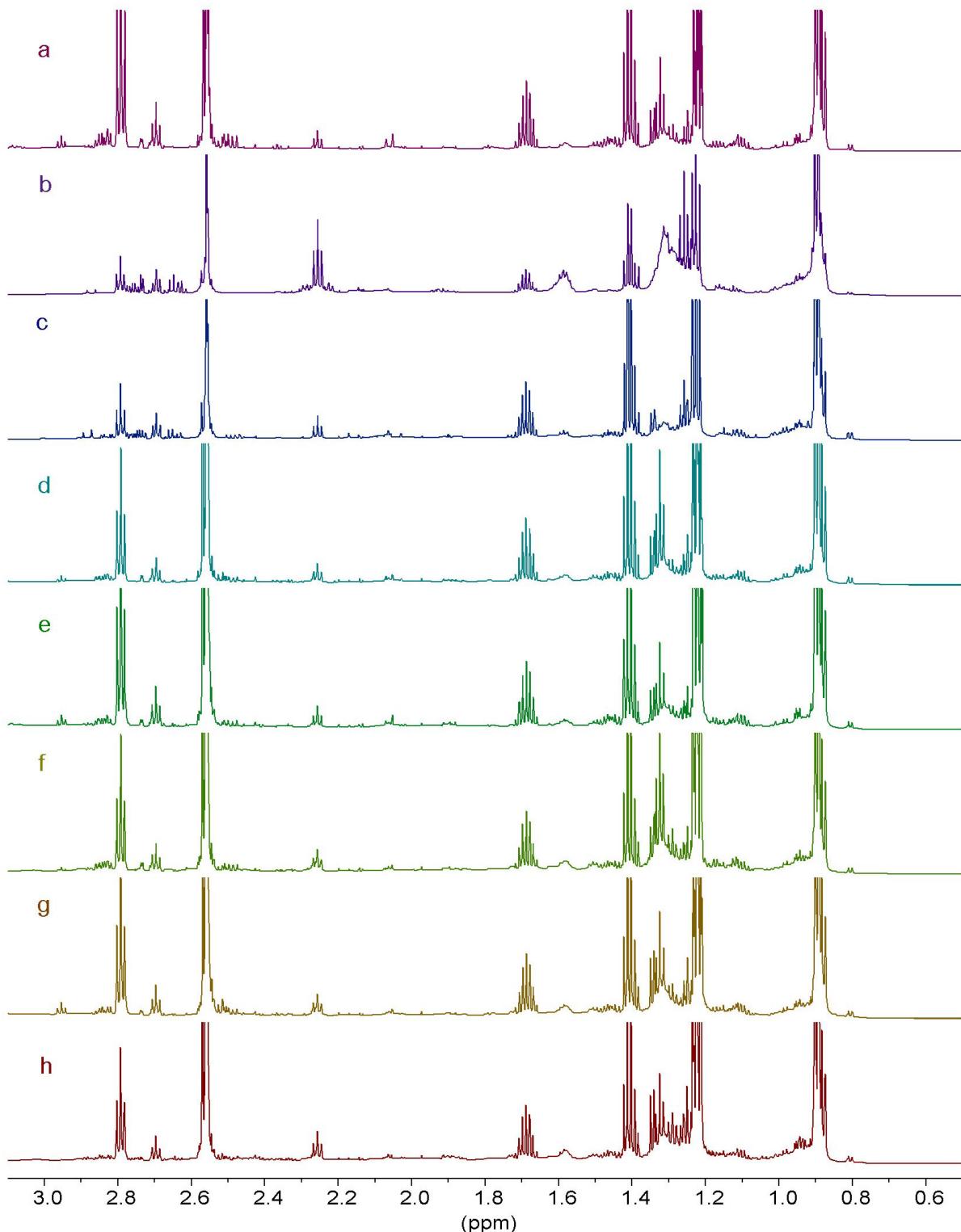
Supplementary Fig. S1: ^1H -NMR expansion spectra (700 MHz, D_2O , 25 °C) from 0.5 to 3.1 ppm of eight wine samples by DA-NMR. Wines: **a**) 'Cabernet Sauvignon', **b**) mixture of 'Chenin Blanc' and 'Colombard', **c**) 'White Zinfandel', **d**) 'Petite Sirah', **e**) 'Merlot', **f**) 'Zinfandel', **g**) 'Nebbiolo' and **h**) 'Barbera'.



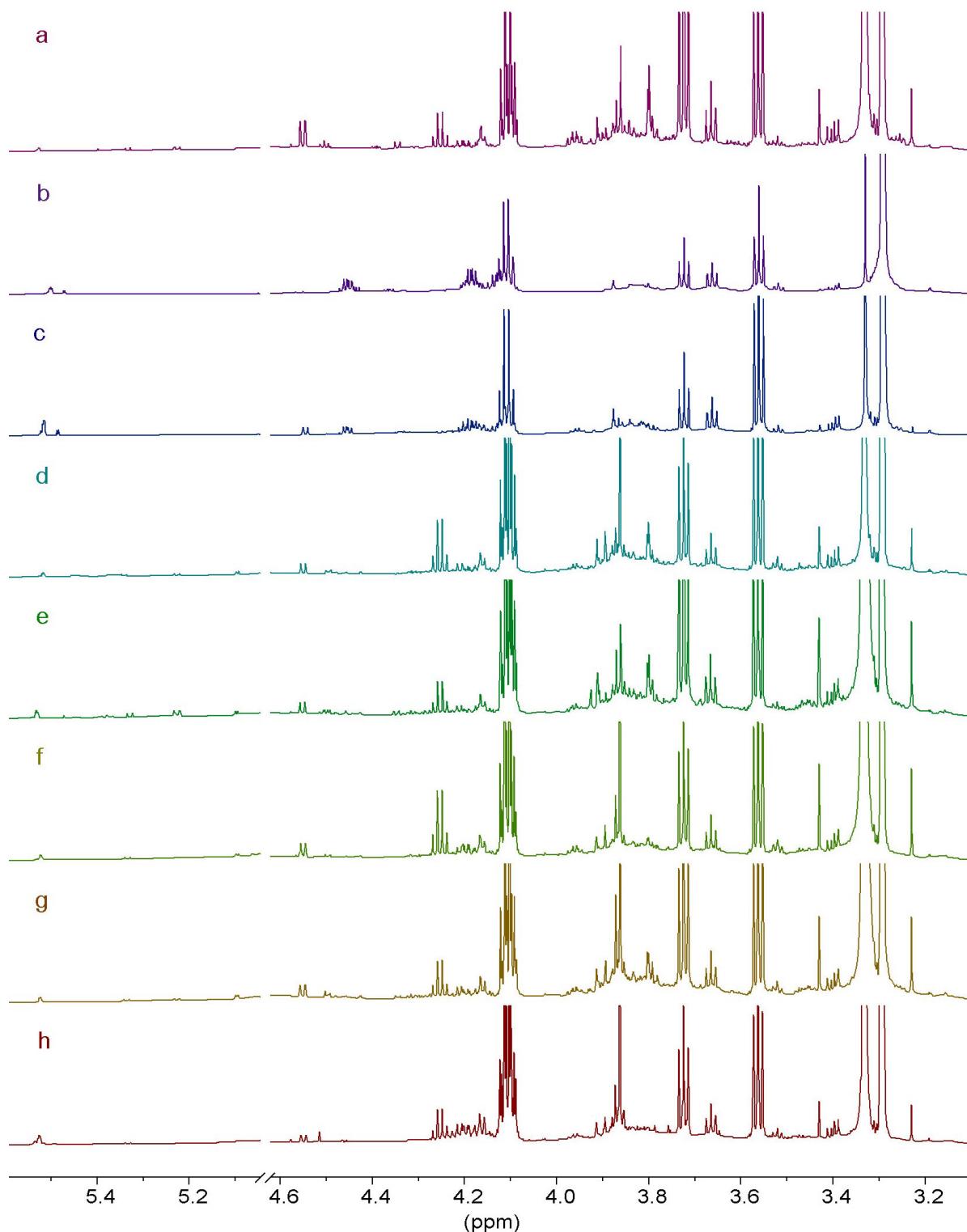
Supplementary Fig. S2: ¹H-NMR expansion spectra (700 MHz, D₂O, 25 °C) from 3.1 to 5.6 ppm of eight wine samples by DA-NMR. Wines: **a**) 'Cabernet Sauvignon', **b**) mixture of 'Chenin Blanc' and 'Colombard', **c**) 'White Zinfandel', **d**) 'Petite Sirah', **e**) 'Merlot', **f**) 'Zinfandel', **g**) 'Nebbiolo' and **h**) 'Barbera'.



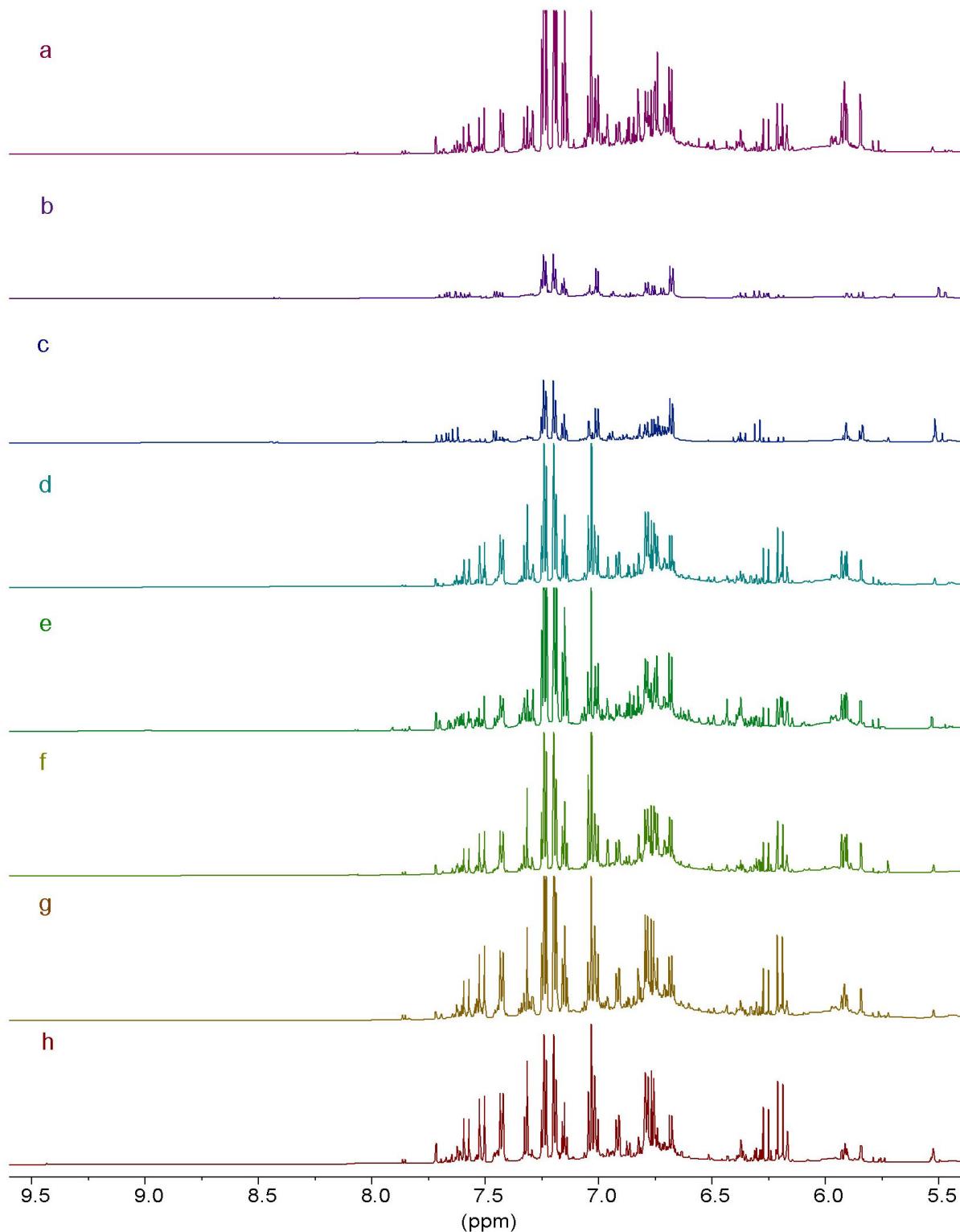
Supplementary Fig. S3: ¹H-NMR expansion spectra (700 MHz, D₂O, 25 °C) from 5.4 to 9.6 ppm of eight wine samples by DA-NMR. The aromatic region was vertical multiplied by factor of thirty. Wines: **a)** 'Cabernet Sauvignon', **b)** mixture of 'Chenin Blanc' and 'Colombard', **c)** 'White Zinfandel', **d)** 'Petite Sirah', **e)** 'Merlot', **f)** 'Zinfandel', **g)** 'Nebbiolo' and **h)** 'Barbera'.



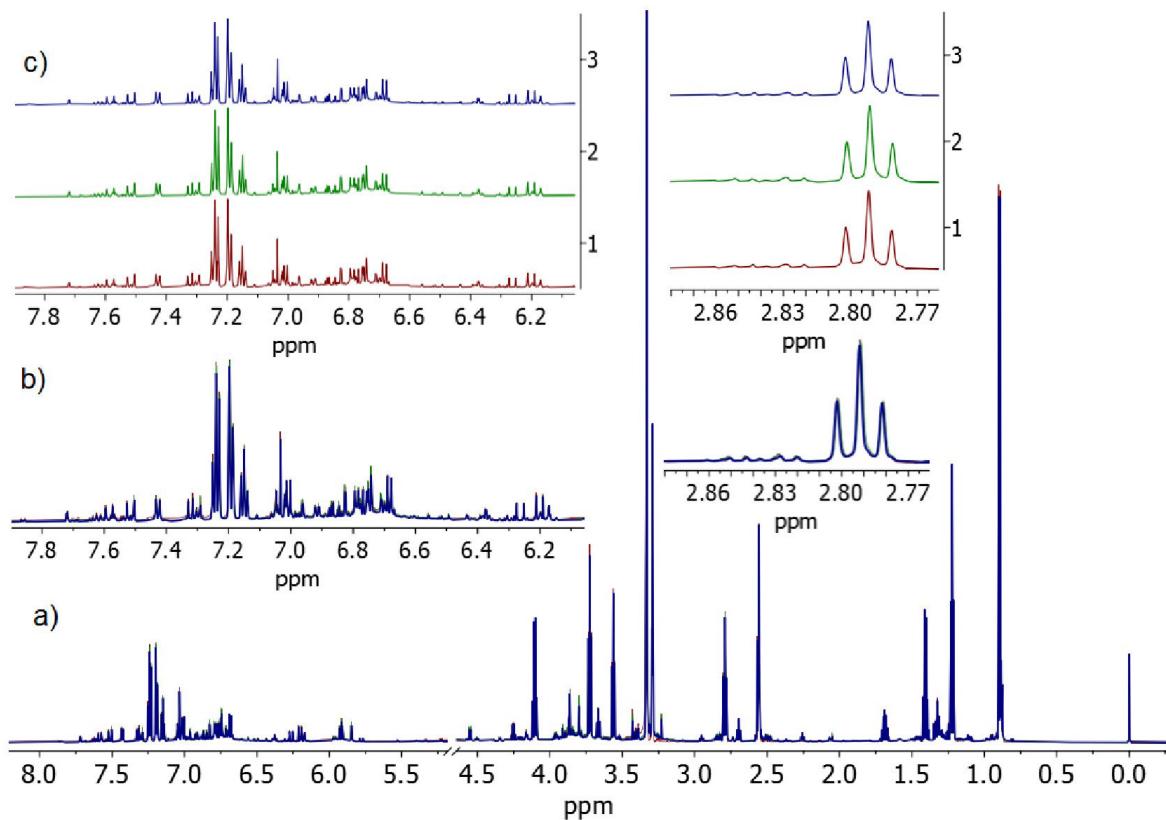
Supplementary Fig. S4: ¹H-NMR expansion spectra (700 MHz, MeOD-*d*₄, 25 °C) from 0.5 to 3.1 ppm of eight wine samples by SPE-NMR. Wines: **a)** 'Cabernet Sauvignon', **b)** mixture of 'Chenin Blanc' and 'Colombard', **c)** 'White Zinfandel', **d)** 'Petite Sirah', **e)** 'Merlot', **f)** 'Zinfandel', **g)** 'Nebbiolo' and **h)** 'Barbera'.



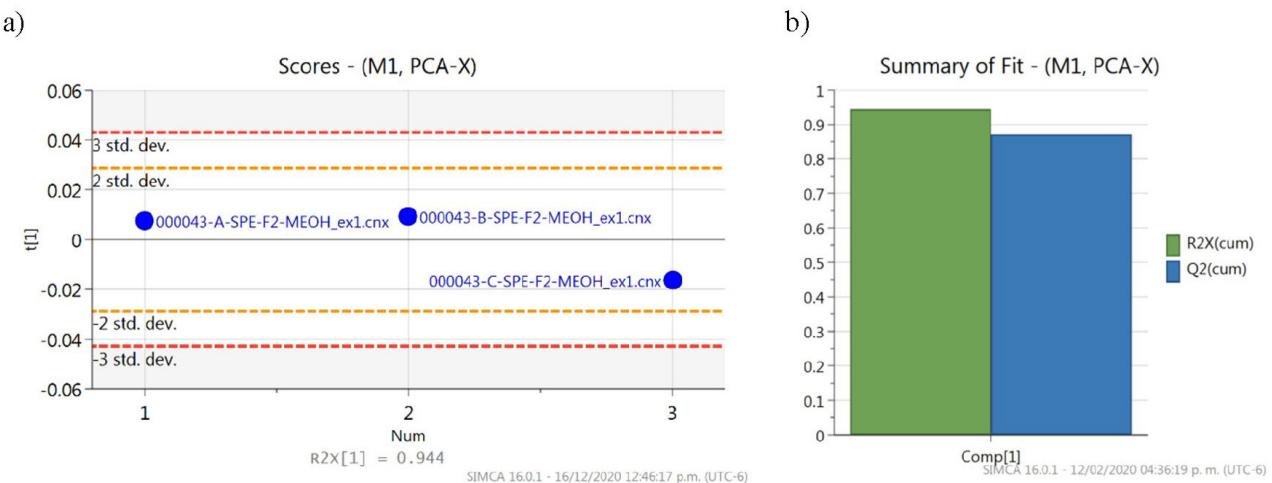
Supplementary Fig. S5: ¹H-NMR expansion spectra (700 MHz, MeOD-*d*₄, 25 °C) from 3.1 to 5.6 ppm of eight wine samples by SPE-NMR. Wines: **a**) 'Cabernet Sauvignon', **b**) mixture of 'Chenin Blanc' and 'Colombard', **c**) 'White Zinfandel', **d**) 'Petite Sirah', **e**) 'Merlot', **f**) 'Zinfandel', **g**) 'Nebbiolo' and **h**) 'Barbera'.



Supplementary Fig. S6: ¹H-NMR expansion spectra (700 MHz, MeOD-*d*₄, 25 °C) from 5.4 to 9.6 ppm of eight wine samples by SPE-NMR. Wines: **a**) 'Cabernet Sauvignon', **b**) mixture of 'Chenin Blanc' and 'Colombard', **c**) 'White Zinfandel', **d**) 'Petite Sirah', **e**) 'Merlot', **f**) 'Zinfandel', **g**) 'Nebbiolo' and **h**) 'Barbera'.

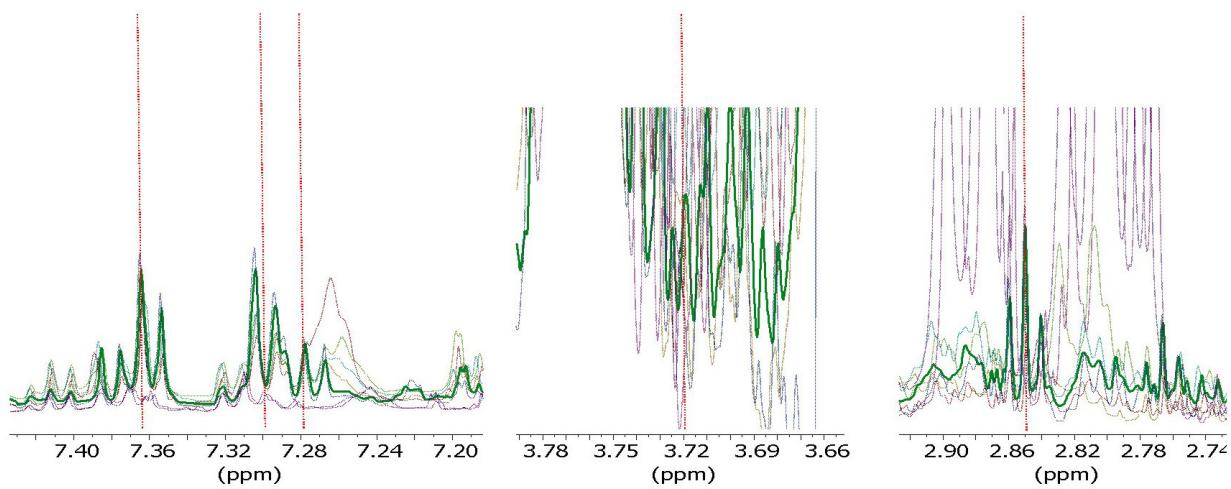


Supplementary Fig. S7: ¹H-RMN spectra (700 MHz, MeOD-*d*₄, 25 °C) of triplicate methanol extracts of a 'Cabernet Sauvignon' wine sample performed to evaluate SPE-NMR repeatability: a) Full overlapped spectra, b) overlapped expansions from 6.05 to 7.9 ppm and 2.76 to 2.88 ppm and c) stacked spectra of same expansions of b).

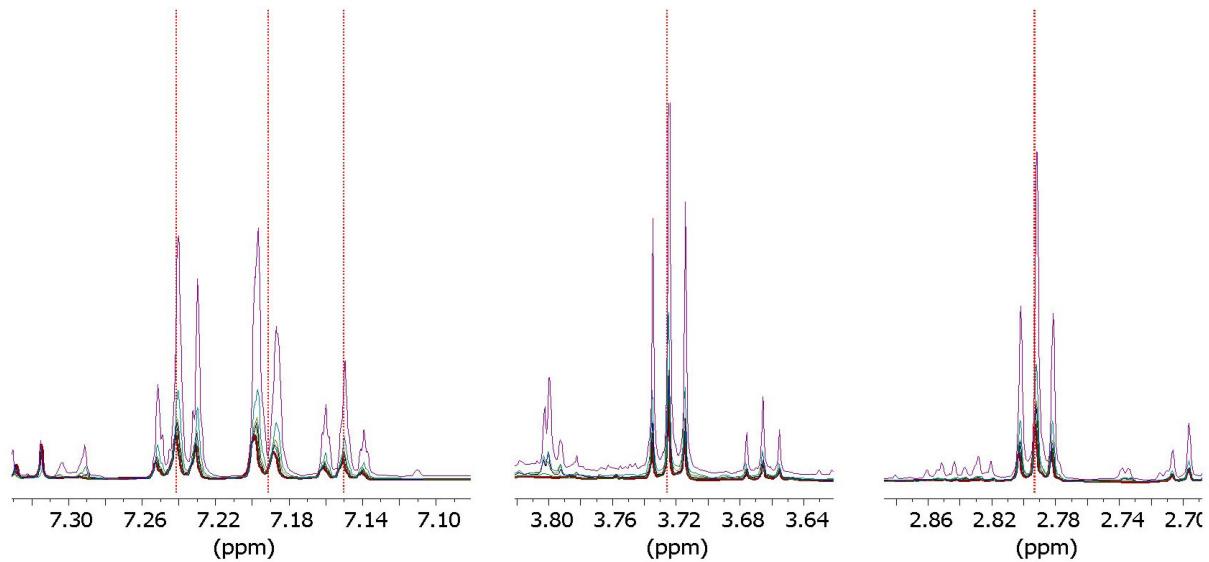


Supplementary Fig. S8: a) Control chart of the first component PC1 that explains 94 % of the data variability of the Fig.S7 spectra and b) cliff plot showing the explained variance of the PCA model with a principal component.

a)

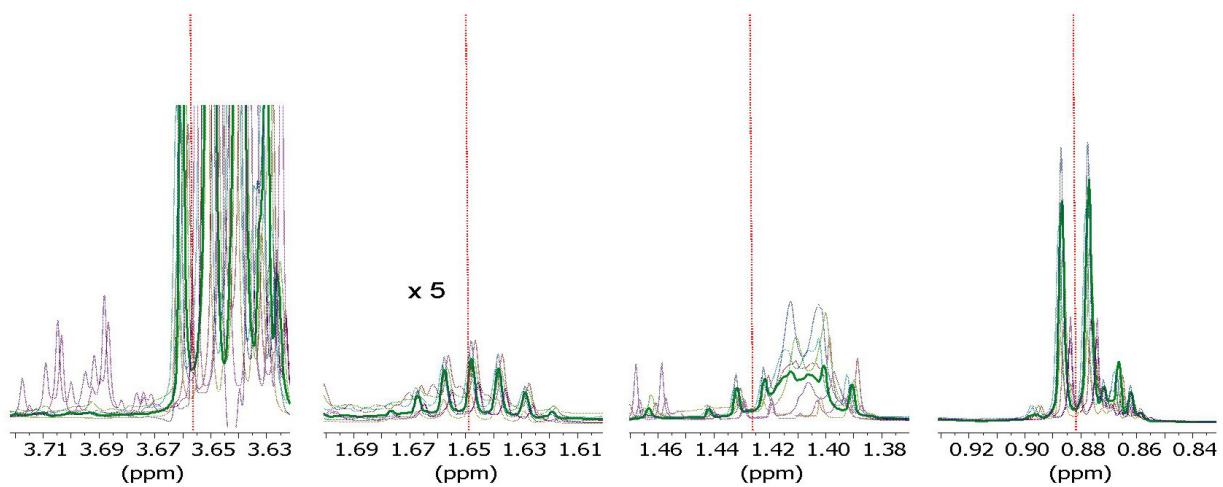


b)

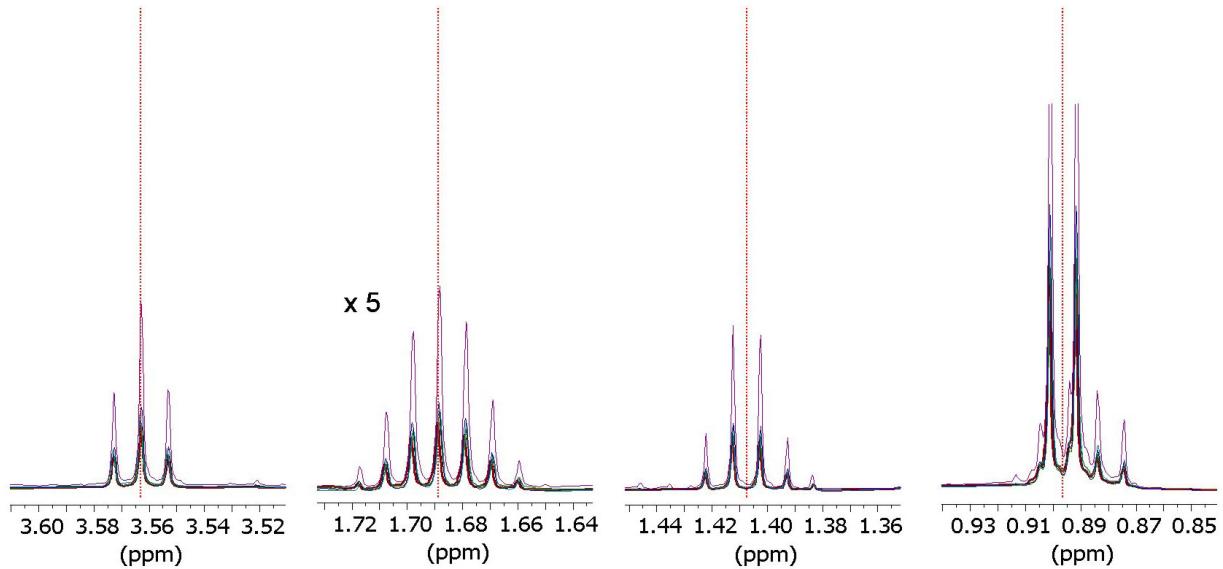


Supplementary Fig. S9: Overlapped ^1H -NMR spectra of phenethyl alcohol signals in the eight wine samples: **a**) DA-NMR and **b**) SPE-NMR.

a)



b)

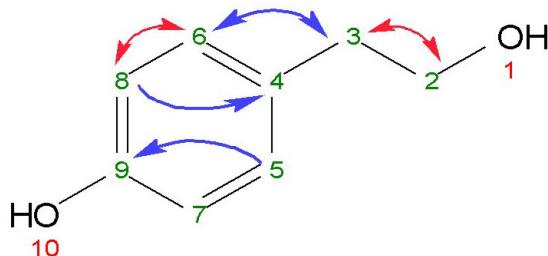


Supplementary Fig. S10: Overlapped ^1H -NMR spectra of isoamyl alcohol signals in the eight wine samples: **a)** DA-NMR and **b)** SPE-NMR. The regions around 1.6-1.7 ppm were multiplied by a factor of five.

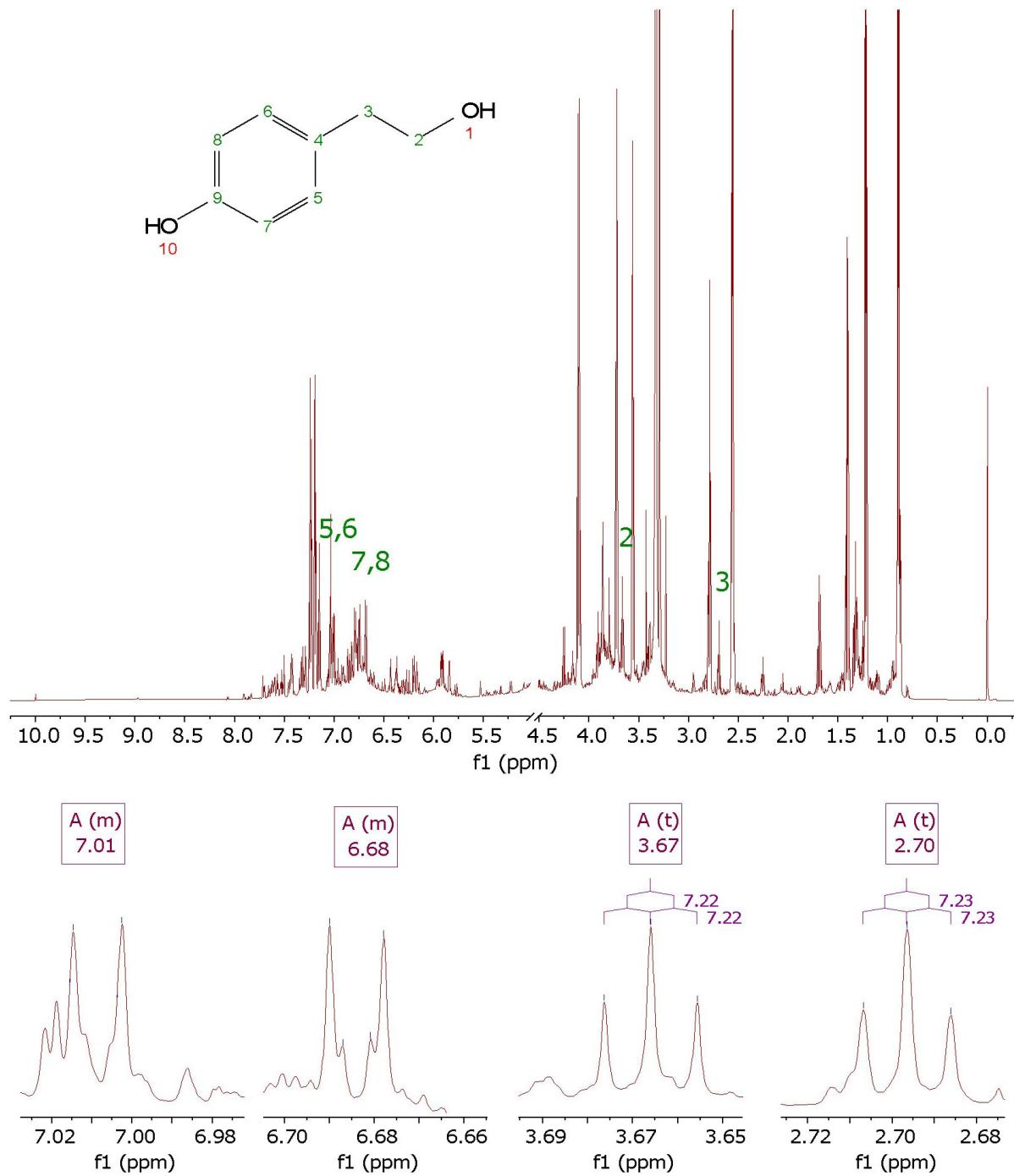
Supplementary work flow S11: Structural elucidation of tyrosol using 1D and 2D NMR spectra in 'Merlot' wine.

The AA'XX' system at $\delta_H = 6.68$ ppm and $\delta_H = 7.01$ ppm (*pseudo-doublets*) suggested a di-substituted phenyl moiety with hydroxyl group in *para* position (**Supplementary Fig. S12**), that could be consistent with tyramine, tyrosol or tyrosine. Interpretation of 2D NMR spectra, i.e., ed-HSQC (edited -Heteronuclear Single Quantum Coherence) (**Supplementary Fig. S15**), HMBC (Heteronuclear Multiple Bond Correlation) (**Supplementary Fig. S16**) and COSY (CORrelation SpectroscopY) (**Supplementary Fig. S14**), allowed identification of tyrosol structure. For instance, the key correlations are:

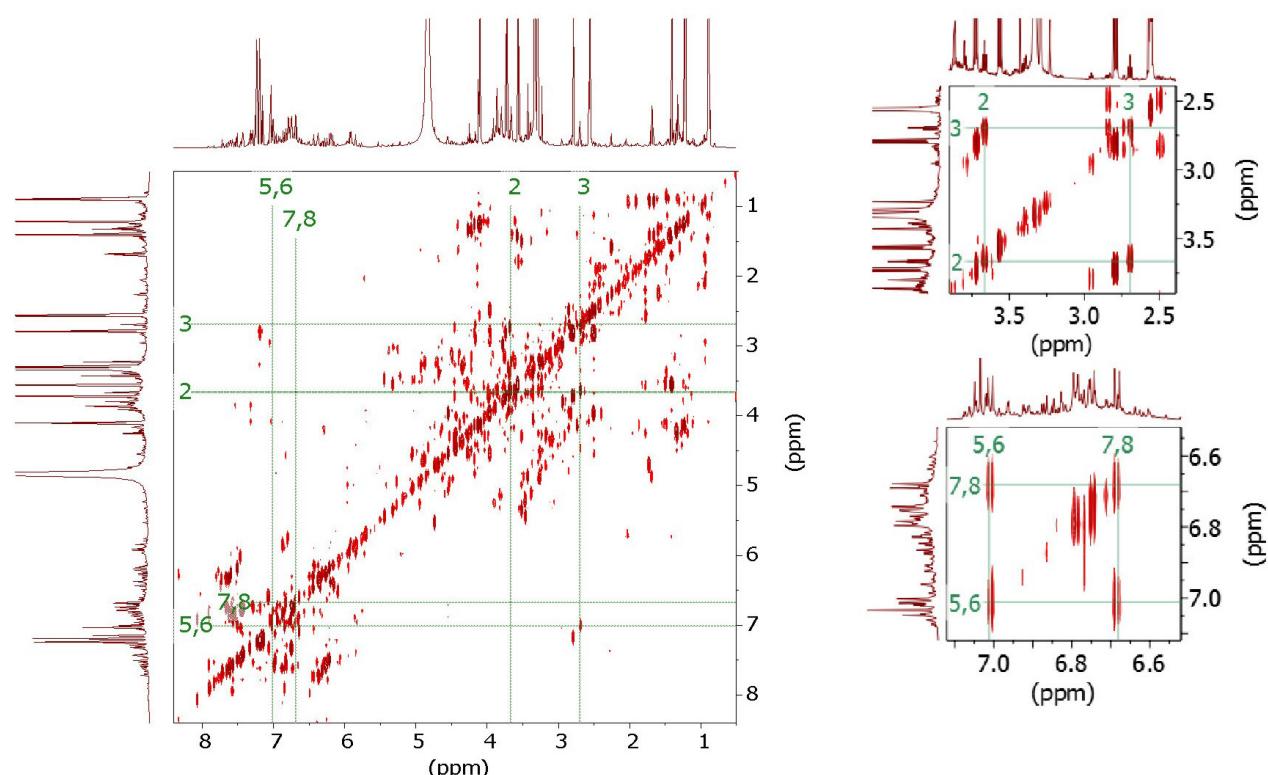
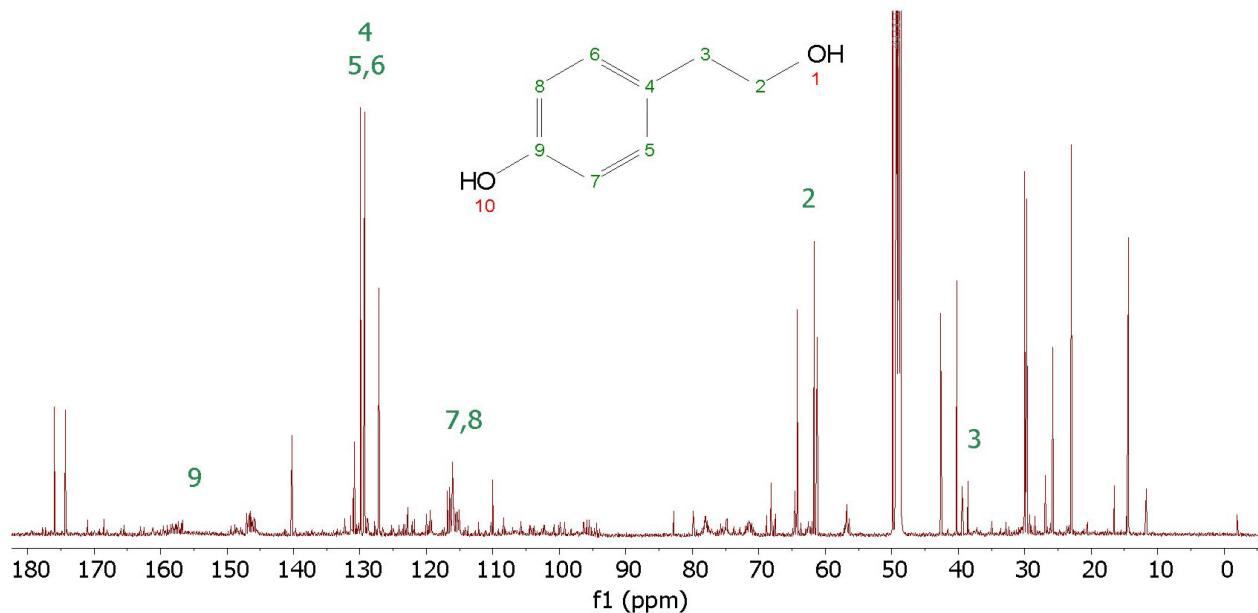
- The correlation between the proton at $\delta_H = 6.68$ ppm (H7,8) and proton at $\delta_H = 7.01$ ppm (H5,6) in the COSY spectra confirm de AA'XX' system of the aromatic ring.
- The quaternary carbons were assigned via $^1H-^{13}C$ HMBC. The correlation from the aromatic protons to the quaternary carbons $\delta_C \sim 155.38$ (C9) and $\delta_C \sim 129.7$ (C4), indicated that the ring has an OH group attach to C9 and quaternary C4 in *para* position.
- The $^1H-^{13}C$ HMBC correlation from the aromatic proton at $\delta_H = 7.01$ ppm (H5,6) to a carbon $\delta_C \sim 38.12$ (C3) indicates the C3 is connected to C4.
- In the multiplicity-edited HSQC experiments, the amplitude of CH_2 signals is negative (signals in blue color) compared to those of CH and CH_3 groups (signals in red color). Carbon $\delta_C \sim 38.12$ (C3) observed in color blue is a CH_2 group which protons appear at $\delta_H = 2.70$ ppm, $t, J = 7.2$ Hz.
- The correlation between the protons at $\delta_H = 2.70$ ppm (H3) and proton at $\delta_H = 3.67$ ppm (H2) in the COSY spectra, the multiplicity and the integrals in the 1H spectra and the blue color in the edited-HSQC confirm that are two connected CH_2 groups, discarding the tyrosine.
- The chemical shift of the last CH_2 ($\delta_H = 3.67$ ppm, $\delta_c \sim 63.36$ ppm) is the expected to be near a OH group and discard the tyramine where a CH_2 near a NH_2 is expected around 42 ppm.

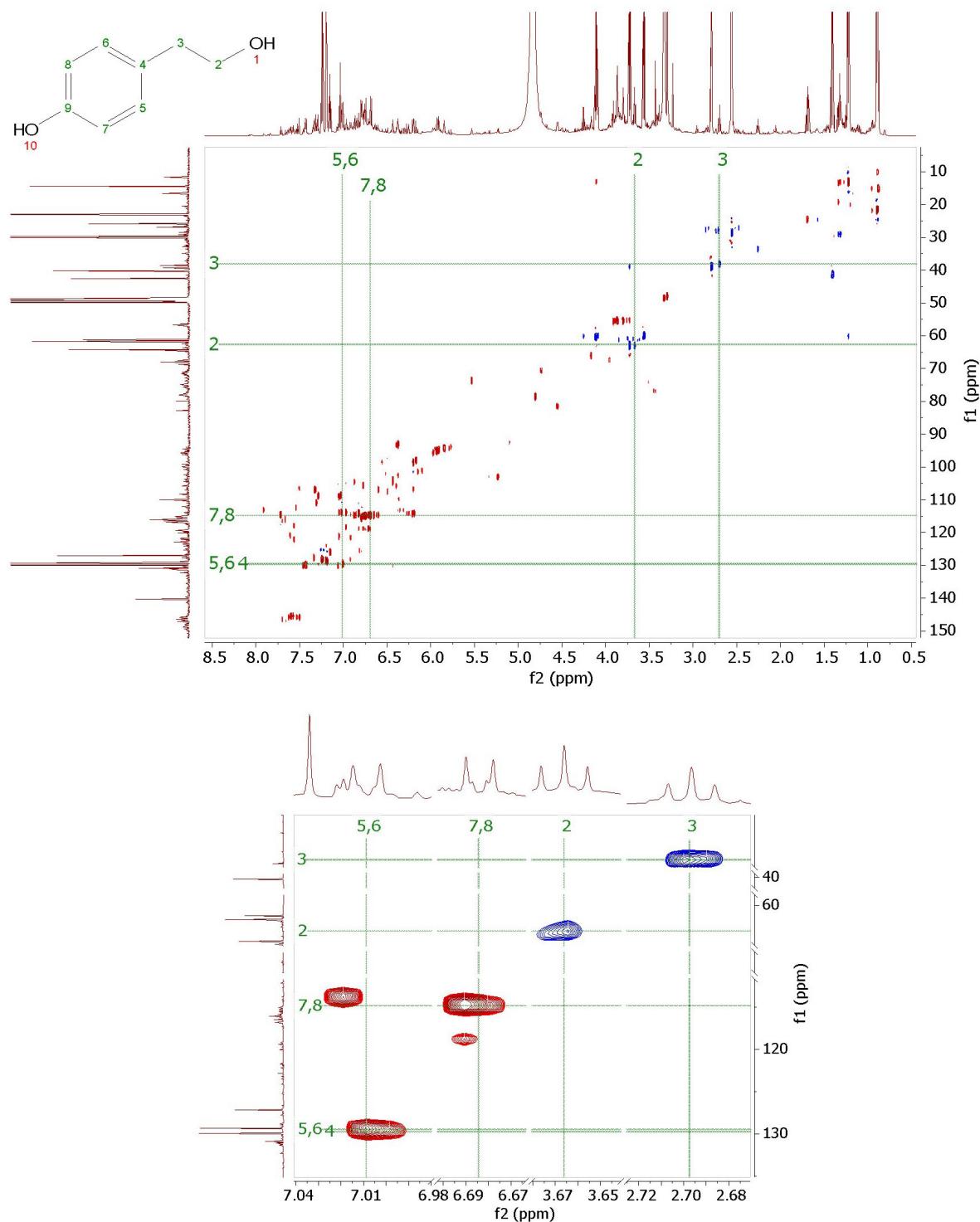


Selected HMBC (blue arrows) and COSY (red arrows) key correlations of tyrosol.
(The numbering used is only for assignment purposes).

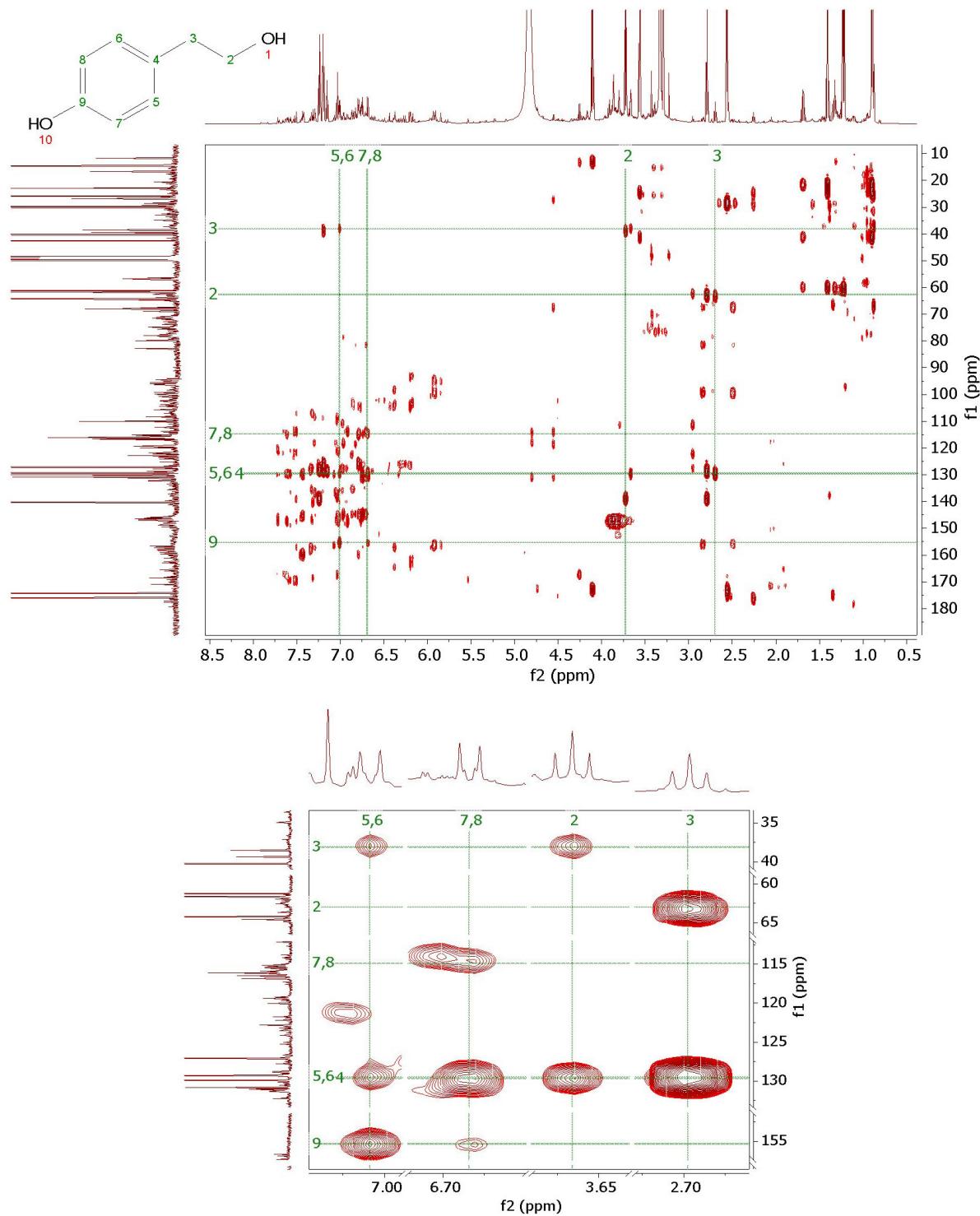


Supplementary Fig. S12: ¹H-NMR spectrum (700 MHz, MeOD-d₄, 25 °C) of Merlot wine extracted by SPENMR. Signal assignments of tyrosol Top full spectrum. Bottom expanded signals of tyrosol.





Supplementary Fig. S15: HSQC spectrum of wine extracted by SPE-NMR (top) full spectrum and (bottom) expanded regions of tyrosol signals.



Supplementary Fig. S16: HMBC spectrum of wine extracted by SPE-NMR (top) full spectrum and (bottom) expanded regions of tyrosol signals.