# Determination of the photosynthetic capacity of grapevine leaves

by

#### H. DURING

Bundesanstalt für Züchtungsforschung im Wein- und Gartenbau, Institut für Rebenzüchtung Geilweilerhof, Siebeldingen, BR Deutschland

## Bestimmung der Photosynthesekapazität in Rebblättern

Zusammenfassung: An Blättern ausreichend bewässerter Topfreben im Freiland wurden vormittags und nachmittags Messungen der stomatären Leitfähigkeit und der Photosynthese durchgeführt, wobei nachmittags stets eine geringere Photosynthesekapazität (maximale Photosyntheserate bei Licht- und  $\mathrm{CO_2}$ -Sättigung) ermittelt wurde. Dieser Befund war mit einer höheren Empfindlichkeit der Stomata gegenüber  $\mathrm{CO_2}$  am Nachmittag verbunden: 0,0016 µbar $^{-1}$  am Vormittag, 0,0046 µbar $^{-1}$  am Nachmittag. Eine Bestimmung der Photosynthesekapazität am Morgen bei 1–2 %  $\mathrm{O_2}$  (Hemmung der Lichtatmung) ergab maximale Photosynthesewerte bereits bei geringeren  $\mathrm{CO_2}$ -Partialdrücken. Dies führte dazu, daß die Photosynthesekapazität bei maximaler stomatärer Leitfähigkeit bestimmt werden konnte. Wurde von den so ermittelten Photosynthesekapazitätswerten der Anteil der Lichtatmung in Abzug gebracht, erreichte die Photosynthesekapazität Werte, die denen bei 21 %  $\mathrm{O_2}$  und höheren  $\mathrm{CO_2}$ -Partialdrücken entsprachen. Dies läßt den Schluß zu, daß in den vorliegenden Untersuchungen am Morgen die Photosyntheserate durch Stomata nicht vermindert war. Zur Verhinderung von Photosyntheseeinbußen infolge Lichthemmung wird vorgeschlagen, die Dauer des Einsatzes von Starklicht zu minimieren und die Lichtintensität auf Werte zu reduzieren, bei denen eine Lichtsättigung gerade erreicht ist.

Key words: leaf, stoma, light, photosynthesis, photosynthetic capacity, carboxylation efficiency, stomatal conductance, respiration, photorespiration, method.

#### Introduction

In numerous experiments the photosynthetic rate of leaves of grapevines has been determined at optimum humidity and temperature conditions and light saturation, e.g. to compare varieties or to study effects of manipulations (e.g. KRIEDEMANN and LENZ 1972; CHAVES et al. 1987; DURING 1988; ROPER and WILLIAMS 1989).

This 'maximum photosynthetic activity' of Riesling leaves varied over a large scale  $(3.2-13.0~\mu\text{mol}~\text{CO}_2\cdot\text{m}^{-2}\cdot\text{s}^{-1})$  among several publications (SCHULTZ 1989). At least in part this variation is due to stomatal conductance, which in grapevine leaves is closely correlated to the rate of photosynthetic  $\text{CO}_2$  assimilation and which fully accounts for the inhibition of  $\text{CO}_2$  assimilation under conditions of moderate water stress by limiting the  $\text{CO}_2$  supply to the mesophyll (Downton et al. 1988 b; During 1987, 1988, 1990). In order to overcome stomatal limitation of  $\text{CO}_2$  supply Lange et al. (1986) determined the 'photosynthetic capacity', i.e. the maximal net photosynthetic  $\text{CO}_2$  assimilation under saturating light conditions and at saturating  $\text{CO}_2$  partial pressure. Saturation of photosynthetic carboxylation reactions with  $\text{CO}_2$  appears to be a promising method to eliminate stomatal influences. However, increasing  $\text{CO}_2$  partial pressures may have a stomata closing effect, which in grapevine leaves has been shown to occur in a non-uniform manner (Raschke 1975; Downton et al. 1988 a, b). The net effect of this non-uniform stomatal behaviour (some groups of stomata close while others remain open) is a reduction in functional leaf area which can lead to erroneous determinations of the

50 H. DURING

photosynthetic capacity. In addition, in glasshouse-grown vines non-stomatal limitations of photosynthetic capacity are reported after prolonged high light conditions (CORREIA *et al.* 1990).

The aim of this paper is to examine the responses of stomatal conductance and  $\rm CO_2$  assimilation to increasing ambient  $\rm CO_2$  partial pressures during different periods of the day and to propose experimental conditions where the determination of the photosynthetic capacity is not disturbed by stomatal or non-stomatal interference.

### Material and methods

3-year-old container-grown Riesling vines grafted to rootstock Kober 5 BB were cultivated outside the glasshouse. Plants were fully irrigated in the evening to keep the leaf water potential above -8 bar during the day. Gas exchange measurements were performed using a Miniküvetten-System (H. Walz, D-8521 Effeltrich, FRG) which has been described earlier (Lange et al. 1986; DURING 1988). At the end of June 1990 the responses of CO<sub>2</sub> assimilation (A) and stomatal conductance (g) to various ambient CO<sub>2</sub> partial pressures were determined in the morning from 9 to 11 a.m. and in the afternoon from 1 to 3 p.m.. Parts of fully expanded but not senescent leaves in the middle part of the vertically trained canes (length: 1.80 m) were inserted into the gas exchange chamber. The cuvette temperature was kept constant at 22 °C, incoming air humidity was controlled at a dew point of 14 °C, thereby producing a relative humidity of about 65 %. Saturating light (1180  $\mu$ mol quanta · m<sup>-2</sup> · s<sup>-1</sup>) was provided for the enclosed part of the leaf with a 12 V/75 W projector lamp and gray filters. CO<sub>2</sub> partial pressure was controlled by a mass-flow-metered gas mixing system and an infrared gas analyser (Binos 1.4 B, Leybold-Heraeus, Hanau, FRG) in the range of 50-2000 μbar. Data obtained for differentials in CO<sub>2</sub> and water vapour, light, temperature and air humidity were stored by a data acquisition system (DES, H. Walz) and computed according to VON CAEMMERER and FARQUHAR (1981). Photorespiration was calculated according to SHARKEY (1988). Stomatal sensitivity to CO<sub>2</sub> at 350 µbar was determined as  $\Delta g/g \cdot \Delta c_i$ .

A, g,  $c_i$  (intercellular  $CO_2$  partial pressure) and CE (carboxylation efficiency) values which were calculated from data obtained in the afternoon under the assumption of homogeneous stomatal aperture on the leaf surface will be marked with an asterisk (\*) in the following text.

#### Results and discussion

Fig. 1 shows the responses of stomatal conductance for  $CO_2$  ( $g_{CO_2}$ ) and  $CO_2$  assimilation (A) to changes in ambient  $CO_2$  partial pressure ( $c_a$ ) under conditions of saturating light intensity and constant air humidity and temperature. Measurements in the morning indicate an increase of  $g_{CO_2}$  up to a maximum at  $c_a = 450$  µbar and a subsequent decrease to about 40 % of the maximum at  $c_a = 2000$  µbar. The photosynthetic activity plotted against  $c_a$  obviously results in a saturation-type curve, the initial slope, representing the carboxylation efficiency, being almost linear.  $CO_2$  saturation of the  $CO_2$  fixation sites in the chloroplasts is reached at about  $c_a = 1400$  µbar which corresponds to a calculated intercellular  $CO_2$  partial pressure of about 1180 µbar and a  $CO_2$  assimilation rate of 16.4 µmol  $\cdot$  m<sup>-2</sup>  $\cdot$  s<sup>-1</sup>. At higher  $c_a$  values the rate of  $CO_2$  assimilation slightly declines. A similar decline of the photosynthetic activity was observed by Stoev and Slavycheva (1982). The leaf was then maintained in the cuvette at 350 µbar  $CO_2$  and at saturating light conditions for 2 h.

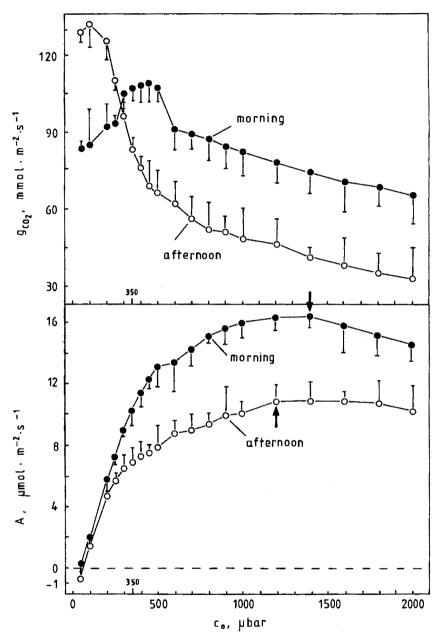


Fig. 1: Stomatal conductance  $(g_{CO_2}^{\bullet})$  and photosynthesis  $(A^{\bullet})$  of leaves of Riesling vines related to ambient  $CO_2$  partial pressure  $(c_a)$ . Measurements were taken in the morning (9-11 a.m.) and the afternoon (1-3 p.m.). Arrows indicate photosynthetic capacity. The vertical bars indicate least significant differences at the 5 % level.

Die stomatäre Leitfähigkeit (g<sub>CO2</sub>\*) und Photosynthese (A\*) von Blättern der Sorte Riesling in Abhängigkeit vom CO<sub>2</sub>-Partialdruck der Außenluft (c<sub>a</sub>). Die Messungen wurden vormittags (9—11 Uhr) und nachmittags (13—15 Uhr) durchgeführt. Die Pfeile verweisen auf die Photosynthesekapazität. Senkrechte Balken: Grenzdifferenzen bei 5 %.

52 H. DÜRING

A repetition of the experiment in the afternoon at the same climatic conditions shows that stomatal conductance\* and  $CO_2$  assimilation\* responses to increasing  $CO_2$  partial pressures were different from those obtained in the morning. At low  $c_a$  values stomatal conductance\* is high (about 130 mmol  $\cdot$  m<sup>-2</sup>  $\cdot$  s<sup>-1</sup>) but declines steadily when  $c_a$  surpasses 100 µbar to values which are about 75 % beyond the initial maximum. At

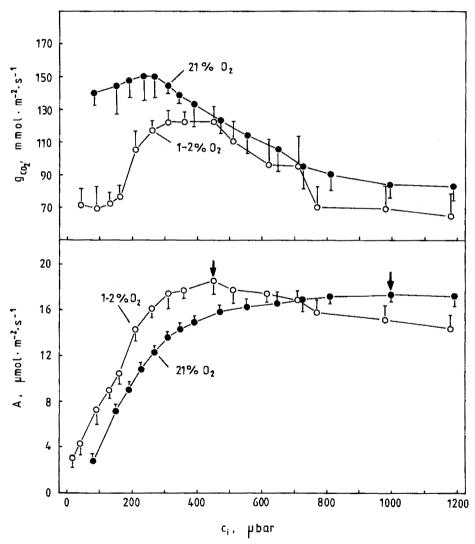


Fig. 2: Stomatal conductance  $(g_{CO_2})$  and photosynthesis (A) of leaves of Riesling vines related to intercellular  $CO_2$  partial pressure  $(c_i)$ . Measurements were taken in the morning (9—11 a.m.) at 21%  $O_2$  or 1—2%  $O_2$ . Arrows indicate photosynthetic capacity. The vertical bars indicate least significant differences at the 5% level.

Die stomatäre Leitfähigkeit ( $g_{CO_2}$ ) und Photosynthese (A) von Blättern der Sorte Riesling in Abhängigkeit vom interzellulären  $CO_2$ -Partialdruck ( $c_i$ ). Die Messungen wurden vormittags (9—11 Uhr) bei 21 %  $O_2$  oder 1—2 %  $O_2$  durchgeführt. Die Pfeile verweisen auf die Photosynthesekapazität. Senkrechte Balken: Grenzdifferenzen bei 5 %.

CO<sub>2</sub> partial pressures  $\geq$  200 µbar photosynthesis\* increases, but to a distinctly lower extent than in the morning. The maximum rate of photosynthesis\* (10.8 µmol·m<sup>-2</sup>·s<sup>-1</sup>) is reached at  $c_a = 1200$  µbar, 2000 µbar CO<sub>2</sub> partial pressure causing a small reduction of CO<sub>2</sub> assimilation\*. It is interesting to note that at low  $c_a$  values (200—250 µbar) the rates of CO<sub>2</sub> assimilation\* in the afternoon are somewhat lower than the corresponding values in the morning, although stomatal conductance\* at 100 to 250 µbar CO<sub>2</sub> is higher in the afternoon. As a consequence, calculated intercellular CO<sub>2</sub> partial pressure\* is higher and the carboxylation efficiency\* is reduced in the afternoon. This may be a hint at mesophyll limitations of photosynthesis.

In another series of experiments (data not shown) stomatal conductance and the photosynthetic activity were also reduced in the morning when  $c_a$  values surpassed 350—450 µbar, while in the afternoon this decrease already started at  $c_a \ge 100$ —250 µbar. Calculation of g\* sensitivity to  $CO_2$  at 350 µbar  $CO_2$  partial pressure indicates higher  $CO_2$  sensitivity in the afternoon (0.0046 µbar<sup>-1</sup>) compared to the morning (0.0016 µbar<sup>-1</sup>); this tendency agrees with results of CORREIA et al. (1990).

Stomata of Vitis vinifera are reported to be particularly sensitive to  $CO_2$  (Loveys and During 1984). Raschke (1975) has pointed out that stomata can be sensitized to  $CO_2$  by raising the abscisic acid (ABA) level in leaves. Our earlier results have shown that ABA increased in vine leaves in the morning and reached a peak of approximately twice the pre-dawn level at midday (Loveys and During 1984; Loveys 1984). Thus, the increased sensitization of stomata to  $CO_2$  in the afternoon may have been mediated by ABA.

It may be argued that even in the morning the determination of photosynthetic capacity may be incorrect, due to stomatal limitation at high CO2 partial pressures. As has been shown earlier under normal atmospheric conditions, photosynthesis of vine leaves is subjected to considerable oxygen inhibition, photorespiration accounting for approximately 20 % of photosynthesis in non-stressed plants (DURING 1988). Determination of photosynthesis at low O2 conditions (1-2 %) will reduce photorespiration to zero without affecting stomatal conductance (Kriedemann and Downton 1981; Downton et al. 1987). In an experiment carried out in the morning with 1—2 % O<sub>2</sub>, the carboxylation efficiency is improved by about 40 % compared to measurements at 21 % O2, as calculated from the data of Fig. 2. The maximum rate of A (18.5  $\mu$ mol  $\cdot$  m<sup>-2</sup>  $\cdot$  s<sup>-1</sup>) is already reached at  $c_i = 450 \mu bar$  (or  $c_a = 600 \mu bar$ ), i.e. at a distinctly lower CO<sub>2</sub> partial pressure compared to the measurement at 21 %  $O_2$  (Fig. 2). At  $c_i = 450 \,\mu bar$  stomatal conductance is still at its maximum (122 mmol  $\cdot$  m<sup>-2</sup>  $\cdot$  s<sup>-1</sup>), only further increases of CO<sub>2</sub> lead to slight decreases of g and A. The maximum value of A can be corrected for photorespiration. Fig. 3 demonstrates that rates of photorespiration per se or related to A decrease at increasing c<sub>i</sub>. At c<sub>i</sub> = 450 µbar (A maximum) photorespiration is diminished to about 10 % of A. As the calculated A capacity  $(18.5-10 \% = 16.7 \mu mol$ .  $m^{-2} \cdot s^{-1}$ ) and the A capacity determined at the same leaf at ambient  $O_2$  conditions  $(17.2 \, \mu \text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1})$  are quite close it is suggested that the distinct decline of stomatal conductance from 150 to 84 mmol  $\cdot$  m<sup>-2</sup>  $\cdot$  s<sup>-1</sup> (Fig. 2) obviously had no limiting effect on photosynthesis.

#### Conclusion

A decline of carboxylation efficiency and photosynthetic capacity at noon or in the afternoon has already been reported for *Arbutus unedo* (BEYSCHLAG 1984; RASCHKE und RESEMANN 1986; DEMMIG-ADAMS et al. 1989) and *Quercus suber* (TENHUNEN et al. 1984). Several hypotheses have been established to explain the phenomenon, including sto-

54 H. DÜRING

matal closure due to decreases of plant water potential or air humidity (SCHULZE 1986), feed-back inhibition due to the accumulation of carbohydrates (AZCON-BIETO 1986) and inhibitory effects after prolonged high light exposure of leaves due to photoinhibition (CORREIA et al. 1990).

The present results indicate that the reductions of carboxylation efficiency and photosynthetic capacity which had been demonstrated for leaves of glasshouse-grown vines (Correia et al. 1990) also occur under outdoor conditions. Although the causes of limited photosynthesis in the afternoon are still rather obscure, it appears that stomatal and non-stomatal effects are involved. There is strong evidence now that, (1) determination of the photosynthetic capacity of grapevines should be restricted to the morning when stomatal sensitivity to  $\mathrm{CO}_2$  obviously is low and (2) prolonged exposure of leaves to high light intensities should be avoided to attain reproducible data of photosynthetic capacity.

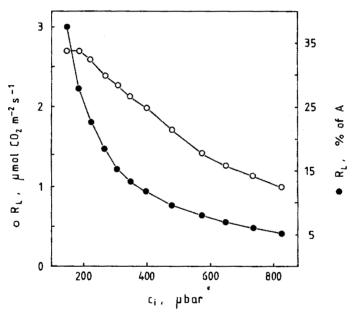


Fig. 3: Photorespiration  $(R_L)$  of leaves of Riesling vines related to the intercellular  $CO_2$  partial pressure  $(c_i)$ . Measurements were taken in the morning (9-11 a.m.) at  $21\% O_2$ .

Die Lichtatmung  $(R_L)$  von Blättern der Sorte Riesling in Abhängigkeit vom interzellulären  $CO_2$ -Partialdruck  $(c_i)$ . Die Messungen wurden vormittags (9-11 Uhr) bei  $21 \% O_2$  durchgeführt.

## Summary

At fully irrigated, container-grown vines (cv. Riesling) kept outdoor, measurements of stomatal conductance and photosynthesis of leaves were performed in the morning and in the afternoon. The results revealed reductions of the photosynthetic capacity (maximal net photosynthesis at saturating light conditions and at saturating  $CO_2$  partial pressure) in the afternoon. This observation was associated with a higher sensitivity of stomata to  $CO_2$  in the afternoon: 0.0016  $\mu$ bar<sup>-1</sup> in the morning, 0.0046  $\mu$ bar<sup>-1</sup> in the afternoon. Due to inhibition of photorespiration determinations of

photosynthetic capacity in the morning at 1-2%  $O_2$  revealed maximal values at lower  $CO_2$  partial pressures and, thereby, at maximal stomatal conductance. These values, corrected for photorespiration, were close to those obtained at ambient  $O_2$  and high  $CO_2$  partial pressures. Thus, it is assumed that in our experiments stomata did not limit the rates of photosynthesis which were determined in the morning. To avoid reduction of photosynthesis by photoinhibition it is proposed to minimize the duration of exposure of leaves to high light and to reduce light intensity to values where photosynthesis begins to reach light saturation.

### Literature

- AZCON-BIETO, J.; 1986: The control of photosynthetic gas exchange by assimilation in wheat. In: MARCELLE, R.; CLIJSTERS, H.; VAN POUCKE, M. (Eds.): Biological Control of Photosynthesis, 231—240. Martinus Nijhof, Dordrecht, the Netherlands.
- Beyschlag, W.; 1984: Photosynthese und Wasserhaushalt von Arbutus unedo L. im Jahreslauf am Freilandstandort in Portugal. Gaswechselmessungen unter natürlichen Bedingungen und experimentelle Faktorenanalyse. Diss., Univ. Würzburg.
- CAEMMERER, S. VON; FARQUHAR, G. D.; 1981: Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves. Planta 153, 376—387.
- Chaves, M. M.; Harley, P. C.; Tenhunen, J. D.; Lange, O. L.; 1987: Gas exchange studies in two Portuguese grapevine cultivars. Physiol. Plant. 70, 639—647.
- CORREIA, M. J.; CHAVES, M. M. C.; PEREIRA, J. S.; 1990: Afternoon depression in photosynthesis in grapevine leaves evidence for a high light stress effect. J. Exp. Bot. 41, 417—426.
- Demmig-Adams, B.; Adams III, W. W.; Winter, K.; Meyer, A.; Schreiber, U.; Pereira, J. S.; Kruger, A.; Czygan, F. C.; Lange, O. L.; 1989: Photochemical efficiency of photosystem II, photon yield of O<sub>2</sub> evolution, photosynthetic capacity and carotenoid composition during the midday depression of net CO<sub>2</sub> uptake in *Arbutus unedo* growing in Portugal. Planta 177, 377—387.
- Downton, W. J. S.; Grant, W. J. R.; Loveys, B. R.; 1987: Diurnal changes in the photosynthesis of field-grown grapevines. New Phytol. 105, 71—80.
- ; LOVEYS, B. R.; GRANT, W. J. R.; 1988 a: Stomatal closure fully accounts for the inhibition of photosynthesis by abscisic acid. New Phytol. 108, 263—266.
- ——;——; 1988 b: Non-uniform stomatal closure induced by water stress cause putative non-stomatal inhibition of photosynthesis. New Phytol. 110, 503—509.
- DURING, H.; 1987: Stomatal responses to alterations of soil and air humidity in grapevines. Vitis 26, 9—18
- ; 1988: CO<sub>2</sub> assimilation and photorespiration of grapevine leaves: Responses to light and drought. Vitis 27, 199—208.
- ——; 1990: Stomatal adaptation of grapevine leaves to water stress. Proc. 5th Intern. Symp. Grape Breeding, 12—16 Sept. 1989, St. Martin/Pfalz, FRG. Vitis, Special Issue, 366—370.
- KRIEDEMANN, P. E.; DOWNTON, W. J. S.; 1981: Photosynthesis. In: PALEG, L. G.; ASPINALL, D. (Eds.): The Physiology and Biochemistry of Drought Resistance in Plants, 283—314. Academic Press, Sydney.
- ; Lenz, F.; 1972: The response of vine leaf photosynthesis to shoot tip excision and stem cincturing. Vitis 11, 193—197.
- Lange, O. L.; Fuhrer, G.; Gebel, J.; 1986: Rapid field determination of photosynthetic capacity of cut spruce twigs (*Picea abies*) at saturating ambient CO<sub>2</sub>. Trees 1, 70—77.
- LOVEYS, B. R.; 1984: Diurnal changes in water relations and abscisic acid in field-grown Vitis vinifera cultivars. III. The influence of xylem-derived abscisic acid on leaf gas exchange. New Phytol. 98, 563—573.
- ; DURING, H.; 1984: Diurnal changes in water relations and abscisic acid in field-grown Vitis
  vinifera cultivars. II. Abscisic acid changes under semi-arid conditions. New Phytol. 97,
  37—47.
- RASCHKE, K.; 1975: Stomatal action. Ann.Rev. Plant Physiol. 26, 309-340.
- ; RESEMANN, A.; 1986: The midday depression of CO<sub>2</sub> assimilation in leaves of Arbutus unedo L.: diurnal changes in photosynthetic capacity related to changes in temperature and humidity. Planta 168, 546—558.

56 H. DURING

- ROPER, T. R.; WILLIAMS, L. E.; 1989: Net CO<sub>2</sub> assimilation and carbohydrate partitioning of grapevine leaves in response to trunk girdling and gibberellic acid application. Plant Physiol. 89, 1136—1140.
- Schultz, H.-R.; 1989: CO<sub>2</sub>-Gaswechsel und Wassertransport von Stark- und Schwachlichttrieben bei Vitis vinifera L. (cv. Riesling) in Abhängigkeit von Klima- und Pflanzenfaktoren. Ansatz eines empirischen Assimilationsmodells. Gesellschaft zur Förderung der FA Geisenheim (Hrsg.), Veröff. der FA Geisenheim Band 5.
- SHARKEY, T. D.; 1988: Estimating the rate of photorespiration in leaves. Physiol. Plant. 73, 147-152.
- Stoev, K.; Slavicheva, T.; 1982: La photosynthèse nette chez la vigne (Vitis vinifera L.) et les facteurs écologiques. Connaiss. Vigne Vin 16, 171—185.
- Tenhunen, J. D.; Lange, O. L.; Gebel, J.; Beyschlag, W.; Weber, J. A.; 1984: Changes in photosynthetic capacity, carboxylation efficiency, and CO<sub>2</sub> compensation point associated with midday stomatal closure and midday depression of net CO<sub>2</sub> exchange of leaves of *Quercus suber*. Planta 162, 193—203.

Received, 27, 2, 1991

Priv. Doz. Dr. H. DURING
Bundesanstalt für Züchtungsforschung
im Wein- und Gartenbau
Institut für Rebenzüchtung Geilweilerhof
D 6741 Siebeldingen
BR Deutschland