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Photochemical and non-photochemical responses of glasshouse-grown grape to combined light and water stress

by

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S u m m a r y : Riesling and Müller-Thurgau vines cultivated in a glasshouse at maximal 400 μ mol quanta m⁻²s⁻¹ were exposed to water logging and subsequently to decreasing soil moisture contents (SMC) until severe drought stress occurred. In the course of the daily applied light treatments, consisting of stepwise increases of light intensity from 0 to 3200 μ mol m⁻²s⁻¹ with intermittent stabilisation phases, the Fv/Fm of dark-adapted and the Fv/F'm ratios of light-adapted leaves were determined by chlorophyll fluorescence measurements. At moderate SMC (21 %) quantum yield of PSII (Y) of Müller-Thurgau leaves decreased when photosynthetic active radiation (PAR) increased beyond 400 μ mol m⁻²s⁻¹. Water stress (water logging and drought) led to a more precipitous decrease, indicating an increased sensitivity of Y to the combined stresses. The deviation of the electron transport rates from (theoretical) maximum quantum yield ("excessive PAR") increased with increasing PAR and was higher in water-stressed than in moderately irrigated vines. Non-photochemical quenching started to increase at relatively low PAR (400 μ mol m⁻²s⁻¹) and reached saturation at 1600 μ mol m⁻²s⁻¹; values were higher in water-stressed vines than in moderately irrigated vines. Riesling and Müller-Thurgau vines showed photoinhibition of photosynthesis at high PAR; water logging and drought intensified this tendency. The extent of photoinhibition was higher in Müller-Thurgau than in Riesling leaves. Even under most severe stress conditions photoinhibition was transient, suggesting complete overnight repair of PSII in both varieties.

K e y w o r d s: grapevine, leaf, light, water stress, photosynthesis, photochemistry, photoinhibition, photosystem II, chlorophyll fluorescence.

Introduction

In many parts of the world, in particular in mediterranean or subtropical climates, grapevines are subjected to periods of low rainfall which are often associated with clear days and high solar radiation. Unless irrigated, grapes may suffer from multiple stresses, i.e. drought, high temperature and high light (OSMOND et al. 1986, VALLADARES and PEARCY 1997). It has been shown earlier that, in contrast to field-grown Riesling vines, CO, assimilation of glasshouse-grown vines exposed to restricted light intensities declined when light intensity increased beyond 800 µmol·m⁻²·s⁻¹. Since this decline could not be ascribed to stomatal closure we assumed that high light intensity might have inactivated the photochemistry of the photosystem II (PSII) reaction centres (photoinhibition) (DÜRING 1988). Evidence for photoinhibition to occur also in field-grown grape was provided by GAMMON and PEARCY (1990), IACONO and SOMMER (1996) and SCHULTZ (1996); in contrast CHAUMONT et al. (1997) demonstrated the absence of photoinhibition in outdoor-grown vines under two different climatic conditions. Moreover, the significance of water stress to enhance photoinhibition is discussed controversially. GAMMON and PEARCY (1990) concluded that the degree of light-dependent decline of PSII fluorescence (Fv/Fm) did not vary with the water status while BJÖRKMAN and Powles (1984) suggested that water stress predisposes the leaves to photoinhibition. Diverging results may be due to differences in plant species, stress intensity or prehistory of plants, e.g. the degree of light adaptation. The aim of this paper was to study the effects of two combined stresses, flooding or drought and increasing light intensity on the quantum yield of PSII under controlled laboratory conditions to better understand the mechanism(s) leading to non-stomatal limitation of photosynthesis in grapes.

Material and methods

P l a n t m a t e r i a l : In March and April 1997 experiments were carried out with 3-year-old potted, glasshouse-grown Müller-Thurgau and Riesling vines. Maximum light intensity at the apical leaves was 400 μ molm⁻²s⁻¹ on clear days. Plants were transferred to the laboratory and kept at 23/17 °C (± 2 °C) (day/night) and a photoperiod of 14 h. Light was provided by Osram HQI lamps, 400 W; 300-400 μ molm⁻²s⁻¹ were measured at the leaf blades selected for experiments. Investigations were started one week after acclimation of vines, which were grown in standard soil ("Fruhstorfer Erde P") containing peat (50 %), granulated volcano-clay (35 %) and bark humus (15 %); they were selected for uniform shoot length (ca. 1.50 m).

Soil humidity and leaf water status: Vines were irrigated daily and supplied with mineral nutrients (Hakaphos grün, Compo, Münster, Germany) twice a week before the experiments were started. At the onset of the experiments they were fully irrigated (100 % water capacity = ca. 50 % soil moisture content (SMC)); thereafter irrigation was withheld for the duration of the experiment (12-14 d). The SMC was determined daily, before and after the light treatments, by a TDR-Trase System (Santa Barbara, California, USA). The leaf water potential was determined at the end of the experiments by the pressure chamber method (SCHOLANDER *et al.* 1965).

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C h l o r o p h y l 1 f l u o r e s c e n c e : Chlorophyll fluorescence was measured at the adaxial part of fully expanded but not senescent leaves with a PAM chlorophyll fluorometer (H. Walz, Effeltrich, Germany). According to SCHREIBER *et al.* (1994) the minimum (Fo) and maximum fluorescence (Fm) following a saturating light pulse (0.7 s, 3500 μ molm⁻²s⁻¹) were used to determine the variable fluorescence (Fv)

$$Fv = Fm - Fo \tag{1}$$

The Fv/Fm ratio of dark-adapted leaves is a measure of the potential (maximum) quantum yield of PSII, the Fv/F'm ratio of light-adapted leaves is an indicator of the actual quantum yield of PSII. The relative electron transport rate (ETR) can be derived from quantum yield of PSII (Y), photosynthetic active radiation (PAR) and a constant (c= 0.42) (SCHREIBER 1997):

$$ETR = Y \cdot PAR \cdot c \tag{2}$$

Non-photochemical fluorescence quenching (NPQ) was calculated according to the Stern-Volmer equation (BJÖRKMAN and DEMMIG-ADAMS 1994):

$$NPQ = Fm/F'm-1$$
 (3)

Light treatments: Each experiment started and ended by darkening a leaf with aluminium foil for 30 min. All treatments/measurements were performed at the same site of a leaf: determination of Fv/Fm (A in Fig. 1), exposure to 20 µmol m⁻² s⁻¹ (Osram L 18 W/77 Fluora) for 20 min, exposure to 400, 800, 1600 and 3200 µmol·m⁻²·s⁻¹ (20 min, fiber illuminator, Type KL 1500, Schott, Mainz, Germany) and determination of Fv/F'm by three saturating light pulses, separated by 30 s, at the end of each light treatment. Between each light treatment the leaf was exposed to 20 µmol·quanta·m⁻²·s⁻¹ (stabilisation phases, according to MAURY et al. 1996). After light treatments and leaf darkening Fv/Fm (B) was determined. The relative rate of photoinhibition (%) was calculated by relating Fv/Fm (B) to the initial Fv/Fm (A) (SCHREIBER 1997). Experiments were repeated 4 times, results in Fig. 3 and 4 were calculated from average values in Fig. 2; results in Fig. 5 were obtained from two consecutive experiments.



Fig. 1: Experimental scheme of light treatments starting with a dark period (30 min) followed by periods of increasing light intensity with stabilisation phases inbetween (PAR, μ mol · m⁻² · s⁻¹) and a final dark period. Fv/Fm - quantum yield of PSII of dark-adapted leaves, Fv/F^{*}m - quantum yield of PSII of light-adapted leaves.

Results

Photochemical and non-photochemical reactions: During their development under glasshouse and laboratory conditions the vines used in our experiments were kept under restricted light conditions (maximum light intensity (PAR) ca. 1/5 of full sunlight). Experiments were started with fully irrigated vines. By withholding irrigation soil moisture content (SMC) decreased slowly to 5.5 % (predawn leaf water potential = -0.60 MPa). Fig. 2 illustrates the relative quantum yield of PSII (Y) at various PARs and high, medium and low SMCs. Irrespective of SMC the initial Y-values of dark-adapted leaves were always close to 0.80 (=100 %) indicating fully open PSII centres and complete overnight recovery from previous high light stress during the experiment (SCHREIBER 1997). At medium SMC (21.1 %) Y started to decrease at PAR >400 μ mol^{-m²s⁻¹}. At high and low SMC increasing PAR led to a precipitous lowering of Y compared to medium SMC. Fig. 3 shows the relative electron transport rate (ETR) in leaves at the three different SMCs as a function of PAR. At high and low SMC



Fig. 2: Quantum yield of PSII (Fv/F'm) of Müller-Thurgau leaves exposed to increasing light intensities at three different soil moisture contents (%). Bars denote 95 % confidence limits.



Fig. 3: Relative electron transport rate (ETR) as a function of PAR in leaves of Müller-Thurgau at medium (21.1 %), high (49.1 %) and low (5.5 %) soil moisture content. Line "a" is the (theoretical) optimum quantum yield line, 'excessive PAR', e.g. at medium soil moisture content, can be calculated by relating A - B to A' - B' (for details: Material and methods).

ETRs were reduced compared to medium SMC; e.g. at 1600 µmol m⁻² s⁻¹ ETR was reduced by 14 % (high SMC) and 29 % (low SMC). At this light intensity 'excessive PAR' (SCHREIBER et al. 1994), calculated from experimental light response curves and the maximum quantum yield line "a" in Fig. 3, was highest at 5.5 % SMC (960 µmol m⁻² s⁻¹) followed by 49.1 % SMC (816 µmol·m⁻²·s⁻¹) while at 21.1 % SMC "excessive PAR" was 640 µmol m⁻² s⁻¹. It is interesting to note that even under favorable soil moisture conditions the capacity of photosynthetic electron transport is limited to the light intensity the vines were adapted to before the onset of experiments. Responses of non-photochemical quenching (NPQ) to increasing PAR at the three soil moisture levels are shown in Fig. 4. Even at low PAR drought and flooding led to distinctly higher NPQ values compared to medium SMC. Independent of SMC NPQ saturation was observed at PAR $>1600 \,\mu\text{mol}\,\text{m}^{-2}\,\text{s}^{-1}$ indicating that the maximum capacity was reached and that leaves were unable to further counteract the increase of excess excitation energy (BJÖRKMAN and DEMMIG-ADAMS 1994).



PAR, mmol quanta m⁻² s⁻¹

Fig. 4: Non-photochemical quenching (NPQ) related to PAR at 5.5, 49.1 and 21.1 % soil moisture content. Variety: Müller-Thurgau.

Evidence for photoinhibition: According to Powles (1984), DEMMIG-ADAMS and ADAMS III (1992), LONG et al. (1994) and TREBST (1994) light intensities exceeding the photosynthetic demand can cause a slowly reversible retardation of photosynthesis, independent of any developmental change, due to a damage of PSII, the so-called photoinhibition of photosynthesis. SCHREIBER (1997) proposed to determine photoinhibition by the rate of recovery of Fv/Fm in a dark period after high light treatment. Fig. 5 shows the relative quantum yield of PSII (B/A ratio, Fig. 1) of Riesling and Müller-Thurgau leaves which were exposed daily to the light treatment during the decline of SMC from ca. 50 % to ca. 4 % corresponding to a decrease of the predawn leaf water potential from zero to -0.7 MPa. Obviously under flooding (SMC >40 %) and drought conditions (SMC <15 %) the relative quantum yield of PSII (Y) was reduced as compared to the medium range of SMC. Except for extreme drought (SMC <5 %) Y values of Riesling were



Fig. 5: Light- and water stress-induced photoinhibition of photosynthesis of Riesling and Müller-Thurgau vines. The relative quantum yield of PSII was calculated by relating Fv/Fm after the light treatment (B) to Fv/Fm before the light treatment (A) (for details:
Fig. 1). The correlation coefficient (r²) for Riesling is 0.86, for Müller-Thurgau 0.69.

always higher than those of Müller-Thurgau. It is interesting to note that even under favorable SMC (ca. 20-35 %) most Y values of Riesling and Müller-Thurgau were depressed. This indicates that the PSII function is partly inactivated even at favorable SMC due to excessive light, leaves of Müller-Thurgau being more severely damaged than those of Riesling. In no case reductions of Y were observed the following day at the end of the first dark period, indicating a complete overnight repair of PSII.

Discussion

In this paper we investigated responses of the photosynthetic apparatus of grape leaves to the absorption of increasing levels of visible light. Since we used vines adapted to restricted light (PAR <400 µmol·m⁻²·s⁻¹) the range of PAR which was photosynthetically utilised, was relatively small, in unstressed vines 800 µmol·m⁻²·s⁻¹ leading already to a reduction of quantum yield of PSII and electron transport; water-stressed vines were shown to be still more sensitive. The small range of light utilisation by photosynthesis is also demonstrated by calculating NPQ which increased already at relatively low light intensities reaching saturation at 1600 µmol m⁻²s⁻¹. These observations confirm that "low light leaves" have a limited capacity of photosynthetic electron transport and energy conversion but also a limited capacity for photoprotective responses such as energy dissipation. In contrast, sun-adapted, outdoor-grown grapes had shown no depression of photosynthesis at increasing PAR even when exposed to 2250 µmol·m⁻²·s⁻¹ (DÜRING 1988). An exposure of our glasshouse-grown vines to light intensities exceeding maximum solar radiation caused "photoinhibition" according to the definition of SCHREIBER (1997). This transient limitation of quantum yield of PSII was modified by

two factors, soil water and genotype. Obviously high light combined with soil water stress (water logging or drought) had an additive effect, i.e. both stresses depressed quantum yield of PSII more than high light treatments at moderate soil water conditions. The transient character of quantum yield limitation is demonstrated by the complete overnight repair of PSII, indicating that the "PSII repair cycle" (migration of light-damaged PSII centres out of the granal thylakoids and migration of undamaged PSII centres into the stromal lamellae) might have been involved (for details: DEMMIG-ADAMS and ADAMS III 1992, ARO et al. 1993, LONG et al. 1994). The variety-specific reactions of photoinhibition may be associated with leaf structure since Riesling leaves showing less photoinhibition are thicker and resemble more to a sunadapted leaf type than leaves of Müller-Thurgau. Moreover, Riesling leaf tissue has been shown to be more drought resistant due to its higher elasticity compared to Müller-Thurgau (DÜRING 1986) which might have contributed to mitigate the stress response in Riesling leaves in the present experiments as well. Light-adapted, field-grown vines were shown recently not to be affected by photoinhibition (CHAUMONT et al. 1997); however, from the present results it can be deduced that photosynthesis of shade-adapted leaves in the inner part of a canopy is reduced by high solar radiation, e.g. after summer pruning or by sunflecks (KRIEDEMANN et al. 1973).

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References

- ARO, E. M.; VIRGIN, I.; ANDERSSON, B.; 1993: Photoinhibition of photosystem II. Inactivation, protein damage and turnover. Biochim., Biophys. Acta 1143, 113-134.
- BJÖRKMAN, O.; DEMMIG-ADAMS, B.; 1994: Regulation of photosynthetic light energy capture, conversion, and dissipation in leaves of higher plants. In: E. D. SCHULZE, M. M. CALDWELL (Eds.): Ecophysiology of Photosynthesis. Ecological Studies 100, 17-47. Springer-Verlag Berlin.
- -; POWLES, S. B.; 1984: Inhibition of photosynthesis reactions under water stress: Interaction with light level. Planta 161, 490-504.
- CHAUMONT, M.; MOROT-GAUDRY, J. F.; FOYER, C. H.; 1995: Effects of photoinhibitory treatment on CO₂ assimilation, the quantum yield of CO₂ assimilation, D₁ protein, ascorbate, glutathione and xanthophyll contents and the electron transport rate in vine leaves. Plant Cell Environ. **18**, 1358-1366.

- -; OSORIO, M. L.; CHAVES, M. M.; VANACKER, H.; MOROT-GAUDRY, J. F.; FOYER, C. H.; 1997: The absense of photoinhibition during the mid-morning depression and photosynthesis in *Vitis vinifera* grown in semi-arid and temperate climates. J. Plant Physiol. 150, 743-751.
- DEMMIG-ADAMS, B.; ADAMS III, W. W.; 1992: Photoprotection and other responses of plants to high light stress. Ann. Rev. Plant Physiol. Plant Mol. Biol. 43, 599-626.
- DURING, H.; 1986: Testing for drought tolerance in grapevine scions. Angew. Bot. 60, 103-111.
- -; 1988: CO₂ assimilation and photorespiration of grapevine leaves: Responses to light and drought. Vitis 27, 199-208.
- GAMMON, J. A.; PEARCY, R. W.; 1990: Photoinhibition in Vitis californica: Interactive effects of sunlight, temperature and water status. Plant Cell Environ. 13, 267-275.
- IACONO, F.; SOMMER, K. J.; 1996: Photoinhibition of photosynthesis and photorespiration in *Vitis vinifera* under field conditions: Effects of light climate and leaf position. Austral. J. Grape Wine Res. 2, 10-20.
- KRIEDEMANN, P. E.; TOROKFALVY, E.; SMART, R. E.; 1973: Natural occurrence and photosynthetic utilisation of sunflecks by grapevine leaves. Photosynthetica 7, 18-27.
- LONG, S. P.; HUMPHRIES, S.; FALKOWSKI, P. G.; 1994: Photoinhibition of photosynthesis in nature. Ann. Rev. Plant Physiol. Plant Mol. Biol. 45, 633-662.
- MAURY, P; MOJAYAD, F.; BERGER, M.; PLANCHON, C.; 1996: Photochemical response to drought acclimation in two sunflower genotypes. Physiol. Plant. 98, 57-66.
- OSMOND, C. B.; AUSTIN, M. P.; BERRY, J. A.; BILLINGS, W. D.; BOYER, J. S.; DACEY, W. J. H.; NOBEL, P. S.; SMITH, S. D.; WINNER, W. E.; 1986: Stress physiology and the distribution of plants. Bio. Sci. 37, 38-48.
- PowLES, S.B.; 1984: Photoinhibition of photosynthesis induced by visible light. Ann. Rev. Plant Physiol. 35, 15-44.
- SCHOLANDER, P. F.; HAMMEL, H. T.; BRADSTREET, D.; HEMMINGSEN, E. A.; 1965: Sap pressure in vascular plants. Science 148, 339-346.
- SCHREIBER, U.; 1997: Chorophyllfluoreszenz und photosynthetische Energieumwandlung: Einfache einführende Experimente mit dem TEACHING-PAM Chlorophyll Fluorometer. H. Walz GmbH, Effeltrich/Germany, 79 pp.
- -; BILGER, W.; NEUBAUER, C.; 1994: Chlorophyll fluorescence as a nonintrusive indicator for rapid assessment of *in vivo* photosynthesis. In: E. D. SCHULZE, M. M. CALDWELL (Eds.): Ecophysiology of Photosynthesis. Ecological Studies 100, 49-70. Springer-Verlag Berlin.
- SCHULTZ, H. R.; 1996: Water relations and photosynthetic responses of two grapevine cultivars of different geographical origin during water stress. In: S. PONI, E. PETERLUNGER, F. IACONO, C. INTRIERI (Eds.): Proc. Workshop: Strategies to Optimize Wine Grape Quality, ISHS. Acta Hort. 427, 251-266.
- TREBST, A. 1994: Dynamics in photosystem II structure and function. In: Ecophysiology of photosynthesis. In: E. D. SCHULZE, M. M. CALDWELL (Eds.): Ecophysiology of Photosynthesis. Ecological Studies 100, 3-16. Springer-Verlag Berlin.
- VALLADARES, F.; PEARCY, R. W.; 1997: Interactions between water stress, sun-shade acclimation, heat tolerance and photoinhibition in the sclerophyll *Heteromeles arbutifolia*. Plant Cell Environ. 20, 25-36.

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